Exposure Assessments of Internally Displaced Infants to Aflatoxin M₁ through Breast Milk Feeding, in Damaturu Yobe State

S. Gide¹*, F. A. Warodi¹, S. D. Alegbe² and G. Anas³

¹Desert Research Monitoring and Control Centre, Yobe State University, Damaturu, P.M.B 1144, Nigeria.
²Department of Microbiology, Ahmadu Bello University, Zaria, Nigeria.
³Nigerian Institute for Trypanosomiasis Research, Kebbi State, Nigeria.

ABSTRACT

Aflatoxin M₁ is a biomarker for the detection of breast milk contamination and also a risk factor for early infant’s exposure to the toxin. Exposure assessment of 50 internally displaced infants to aflatoxin M₁ through breast-milk feeding was carried out between (June 2016 to October 2016). High performance liquid chromatography (HPLC) was used to evaluate the level of AFM₁ in mother’s breast milk samples and the infant’s urine samples respectively. Results obtained from the study showed that 96% of the breast milk samples have maximum concentration of 0.0879µg/L with mean value of 0.0582µg/L while, the minimum and maximum excretion concentration of AFM₁ in urine sample of infants was 0.0400µg/L and 0.0651µg/L respectively with mean value of 0.05005µg/L at 88%. The study indicates that the occurrence of AFM₁ in breast milk samples of mothers with the types of food they consumed within 24-48hrs prior to sample collection that predispose the infant’s exposure.

*Corresponding author: E-mail: gidesuleimans@yahoo.com, gidesuleimans@gmail.com;
to AFM$_1$ showed 40% of the women consumed rice and 32% consumed local food (brabisko/biski) and 24% consumed corn meal with statistically significant with P value less than 0.05. From the study the 96% of the infants were exposed to the toxin while 18% of the infants were undetectable, the concentration of AFM$_1$ in all the breast milk samples were found to be higher than the acceptable tolerance level of 0.025µg/L and 0.05µg/L for infants milk by the European Communities and Codex Alimentarius respectively. This possess a concern that internally displaced infants that were on admission in the selected facility where exposed to AFM$_1$.

Keywords: Internally displaced infants; breast milk; Aflatoxin M$_1$; HPLC.

1. INTRODUCTION

Breast feeding is the best and recommended form of infant feeding. Six month is the recommended period of exclusive feeding for infants. Breastfeeding promotes the mother-child relationship and ensures better growth and development of the neonates given them required nutrients, strengthen antibodies and leukocytes [1,2]. Aflatoxins are metabolites of a Fungus of Aspergillus flavus and Aspergillus parasiticus there are various types of Aflatoxin discovered meanwhile the commonest are Aflatoxin B$_1$, B$_2$, G$_1$,G$_2$, M$_1$ and M$_2$ are the secondary metabolites of B$_1$ and B$_2$ respectively [3]. Aflatoxin M$_1$ (AFM$_1$) as a biomarker of Aflatoxin B$_1$ (AFM$_1$) can be used to evaluate aflatoxin exposure through diet for both humans and animals. As a metabolite human exposure to aflatoxin (AFs) occurs through the intake of contaminated agricultural products or the consumption of products from animals that were fed with contaminated feed. This contamination may occur by fungal growth during harvest or improper storage [4]. AFB$_1$ is considering the most toxic among all other forms of Aflatoxin [5].

Toxicological evaluation of aflatoxin over food intake is essential to any risk evaluation and important for determining the relationship observed in humans and exposure to aflatoxin [6,7]. AFB$_1$ is metabolized in animals and the human liver into AFM$_1$ by Cytochrome P450 associated enzymes, and then distributed in serum and excreted into milk and urine. In mammals, AFM$_1$ can be detected in milk within 12 h after the ingestion of AFB$_1$ [8].

The maximum limits for fluid milk and powdered milk should be 0.5 and 5 ng/g, respectively [9]. A technical regulation on the Maximum Tolerated Limit for Mycotoxins in Food has not been defined regarding whether the fluid milk is raw or pasteurized. The European guidelines established the maximum AFM$_1$ levels in both raw milk (0.05 µg/l) and infant formulae (0.025 µg/l) [10]. Based on the potential hazard to infant (zero to 12 months old) health due to carry-over of the aflatoxin biomarker (AFM$_1$) into milk, the aim of this study was to evaluate the exposure of infants to AFM$_1$ through consumption of breast milk.

2. MATERIALS AND METHOD

The study participants were 50 lactating mothers whose babies were on admission in Yobe State Specialist Hospital Damaturu in the north eastern Nigeria.

2.1 Ethical Approval and Parents Consent

Ethical clearance was obtained from the ethical and scientific research committee of the ministry of health. A consent form was given to obtain permission from mothers whose babies were on admission. Only those who gave consent were recruited in the study.

2.2 Sample Size and Study Population

This study was conducted on 50 internally displaced lactating mothers whose infants were on admission in Yobe State Specialist Hospital Damaturu, Yobe State.

2.3 Inclusion Criteria

All infants on admission in the selected health facility and breast fed after 20min before collection of samples also not administered with any antihistamine were included in the study.

2.4 Exclusion Criteria

Lactating mothers who are on admission, infant with Sjogren' syndrome: an autoimmune disease of the salivary gland, mumps, outpatient, and those on radiotherapy were excluded from the study.
2.5 Breast Milk Collection

Breast milk samples were collected by hand expression into glass tubes (50mL); the milk samples were stored at 15°C. Samples were thawed gradually to 4°C and then centrifuged at 10°C at 15rpm. Aflatoxin is water-soluble; hence, the upper creamy layer was discarded and the lower phase was used for the quantitative test [11].

2.6 Structured Questionnaire

Structured questionnaire was design to determine the kinds of food lactating mothers consumed within 24-48hrs that associate them to Aflatoxin exposure.

2.7 High Performance Liquid Chromatography (HPLC), Analysis for Aflatoxin M1

The concentration of AFM1 in breast milk was estimated by HPLC configured with LC-10AD pumps, coupled with tungsten detector RF-10AXL. Excitation and emission wavelengths were set at 350 and 440 nm respectively. The stationary phase was a Gemini Column. The mobile phase was isocratic mixture of methanol/acetoneitrite/water (25:25:50 v/v/v), with a flow rate of 1ml/min and chromatographic run time of 10 min. The values obtained for recoveries and relative standard deviations of the methods of analysis were in agreement with Commission Regulation (EC) No. 401/2006 for methods of analysis of mycotoxins in foodstuffs [12].

An injector with 50 µl loop was used for the determination of AFM1. A calibration curve was constructed for AF M1 using different levels of toxin concentrations with an average of 10 consecutive automated injections of standard solutions of AFM1 purchased from SUPELCO solutions within USA Involving series of dilutions [11].

2.8 Preparation of Urine Samples for High Performance Liquid Chromatography (HPLC)

The extraction and purification of urine samples for AFM1 determination were performed as follows: A 30 ml volume from the urine sample was filtered through a glass microfiber filter paper. Later, 20 ml of filtered extract was transferred to a 50 ml capacity vial and 20 ml of sodium acetate buffer (pH 5.0) was added. The pH of the mixture was measured and corrected to 5.0 using an appropriate volume of a 0.1 M glacial acetic acid solution whenever necessary. The mixture was directly passed through an immunoaffinity column at a flow rate of approximately 1.0–1.5 ml min⁻¹. After adding the mixture the column was washed with 40 ml of ultrapure water. The column was dried by applying positive pressure with a syringe and bound AFM1 was eluted with 2.0 ml of HPLC-methanol which was recovered in a 4 ml vial previously treated with acid. The eluate was evaporated under nitrogen gas and reconstituted with 500 µl of the mobile phase before liquid chromatography analysis. Detection and quantification of sample extracts were performed by high-performance-liquid chromatography (HPLC) with a liquid chromatography system equipped with a LC-10AT Shimadzu pump, a Shimadzu RF-10AXL fluorescence detector (excitation 250 nm and emission 350 nm), an injection volume of 100µl, and a reverse phase column (250- 4.6mm, particle size of 3 µm) and pre column kept at room temperature. The mobile phase consisted of an isocratic mixture of water and acetonitrile at a volume ratio of 75:25 and a flow rate of 1.0 ml min⁻¹.

A calibration curve was prepared using standard AFM1 solutions in mobile phase at concentrations of 0.005, 0.01, 0.02, and 0.03 ng ml⁻¹. The standard obtained (Sigma, St. Louis, MO, USA, and product code 6428, 10 g) as purified crystalline AFM1 was dissolved in HPLC-grade acetonitrile and its concentration was determined by spectrophotometer [13].

2.9 Statistical Analysis of the Data

The Statistical Package for Social Science program (SPSS, Chicago, Illinois, USA) version 21 was used for analysis of data. Data was summarized as mean, standard deviation, for the analysis of the difference in quantitative data between the two groups. The X²-test was used for the analysis of qualitative data. P value was considered significant if it was less than 0.05 Simple linear correlations (Pearson’s correlation for quantitative data) [14].

3. RESULTS

3.1 Analytical Performance

A calibration curve was prepared using standard AFM1 solutions in mobile phase at concentrations of 0.005, 0.01, 0.02, and 0.03 µg/
The limit of detection (LOD) for AFM$_1$ was estimated at 0.01µg/mL and the limit of quantification (LOQ) was 0.05µg/mL. The linearity of the curve was 0.01–0.05 µg/mL. The calibration curve for AFM$_1$ had a linear equation of \( y = 4147.1x - 230.3 \); Fig. 1 gives the calibration curve with a correlation coefficient \( R^2 = 0.993 \) and retention time of 10.0 min.

The percentage occurrence and concentration of AFM$_1$ in breast milk consumed by infants and excretion in their urine as seen in Table 1 shows that 96% of the breast milk consumed by IDP infants have the toxin with minimum concentration of 0.0457µg/L and maximum concentration of 0.0879µg/L and the mean concentration of 0.0582µg/L meanwhile, the minimum and maximum excretion concentrations of AFM$_1$ in urine samples of infants gave 0.0400µg/L and 0.0651µg/L respectively with mean value of 0.05005µg/L at 88% exposed of the toxin from their mother’s breast milk.

Occurrence of AFM$_1$ in breast milk samples of mothers with the types of food the consumed within 24-48hrs prior to sample collection that predispose the ID infants exposure to AFM$_1$ has been illustrated in Table 2; 40% of the women consumed rice and 32% consumed local food (brabisko/biski) and 24% consumed corn meal with statistically significant with \( P \) value less than 0.05.

4. DISCUSSION

Exposure assessment of internally displaced infants to AFM$_1$ through consumption of breast milk in Damaturu was 96% with maximum concentration of 0.0879µg/L of the toxin. The mean value concentration of AFM$_1$ in the breast milk samples was 0.0582µg/L this may be explained that the mothers were exposed to the toxin in their diet. Considerably, another independent screening by Adejumo et al. report that occurrence of AFM$_1$ was 82% in breast milk samples in Ogun state, Nigeria which is lower [11]. While another study by Abdulrazzaq et al. and Sadeghi et al. in United Arab Emirate and Iran respectively reported 99.5% and 98.1% frequencies which is higher [15,16]. The percentage occurrence in this study is higher than that reported in Cameroun and Egypt, 48%, and 56% respectively by Tchana et al. and Polychronaki et al. [17,18]. This could be as a result of differences in the type of food consumed, storage condition of foods and lifestyle of the people as well as level of contamination with the secreting Aspergillus species.

Infant examined are between the age brackets 0-6 month with minimum concentration of 0.04µg/L and maximum concentration of 0.0651µg/L with mean excretion concentration of 0.05005µg/L. This high concentration implies that the infant were exposed to high level of aflatoxin which is indicative from the levels seen in their mothers’ breast milk samples a major food source for the infant which are expected to be breast feed for six months. 88% of the examined children shown exposure to AFM$_1$, which implies that 18% were undetected in the urine samples of the infant which is in line with the study reported in Brazil by Giolo et al. [19].
Table 1. Occurrence and concentration of AFM$_1$ (µg/L) in breast milk consume by infants and excretion in their urine

<table>
<thead>
<tr>
<th>Samples</th>
<th>No. of samples</th>
<th>No. and (%) positive samples</th>
<th>Minimal concentration (µg/L)</th>
<th>Maximal concentration (µg/L)</th>
<th>Mean concentration ±SD (µg/L)</th>
<th>Chi-square value</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Breast milk</td>
<td>50</td>
<td>48(96)</td>
<td>0.0457</td>
<td>0.0879</td>
<td>0.0582 (±0.012)</td>
<td>2.204</td>
<td>0.02</td>
</tr>
<tr>
<td>Infant’s Urine (0-6 months)</td>
<td>50</td>
<td>44(88)</td>
<td>0.0400</td>
<td>0.0651</td>
<td>0.05005(±0.004781)</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

SD. Standard deviation of descriptive quantities

Table 2. Occurrence of AFM$_1$ in breast milk samples of mothers (N=50) in relation to the types of food consumed within 24-48 hours prior to sample collection

<table>
<thead>
<tr>
<th>Consumed foods</th>
<th>No. and (%) of AFM$_1$, positive (Yes) or (No) samples</th>
<th>Chi-square value</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Brabisko/Biski (local food)</td>
<td>16 (32.0) or 0 (0.0)</td>
<td>8.245</td>
<td>0.045</td>
</tr>
<tr>
<td>Rice</td>
<td>20 (40.0) or 0 (0.0)</td>
<td>9.123</td>
<td>0.034</td>
</tr>
<tr>
<td>Corn</td>
<td>12 (24.0) or 2 (4.0)</td>
<td>10.233</td>
<td>0.013</td>
</tr>
</tbody>
</table>

This result can be compared with other researchers that there is a relationship between food intake and urinary excretion of AFM$_1$ [20]. Redzwan et al. reported that there was significant relationship between consumption of AFM$_1$ in milk and excretion in the urine [21].

5. CONCLUSION

The mean concentration of examined breast milk samples was 0.0582µg/L. It should be noted that, the concentration of AFM$_1$ in all the breast milk samples used were higher than the acceptable tolerance level of 0.025µg/L and 0.05µg/L for infants milk by the European Communities and Codex Alimentarius respectively. This possess a concern on internally displaced infants that were on admission in the selected facility where exposed to AFM$_1$. This implies that AFM$_1$ has the potential to be a public health problem in Damaturu.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

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